PCRBIOSYSTEMS
simplifying research

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VeriFi® Mix

Product description

VeriFi® Mix is a convenient high fidelity 2x mix designed for PCR applications where greater sequence accuracy is required, together with improved PCR success rates of long and challenging templates.

VeriFi® Mix contains the engineered and highly processive PCRBIO VeriFi® Polymerase, developed for fast and versatile high fidelity PCR. The enzyme is derived from Pfu DNA polymerase for its 3'-5' exonuclease (proofreading) activity. Several proprietary mutations significantly improve DNA binding and processivity, resulting in shorter extension times (10-30 s/kb), higher yields and the ability to amplify longer and more difficult targets, including eukaryotic genomic templates in excess of 17.5 kb.

The high accuracy and enhanced 3'-5' exonuclease activity of VeriFi® Polymerase result in fidelity that is approximately 100 times higher than Taq DNA polymerase. The enzyme is ideally suited to applications where greater accuracy is required, such as cloning, site-directed mutagenesis and sequencing. PCR products generated with this range of products are blunt ended.

Quality control

PCR Biosystems operates under an ISO 13485 certified Quality Management System. Our products are extensively tested and undergo a comprehensive, multi-step quality control process according to ISO 13485 standards, to ensure optimum performance, consistency and traceability.

Pack size	2x VeriFi® Mix
100 x 50 µL reactions	2 x 1.25 mL
500 x 50 µL reactions	10 x 1.25 mL

Shipping and storage

On arrival the kit should be stored between -30 °C and -20 °C. If stored correctly, the kit will retain full activity until the indicated expiry date. The kit can be stored at 4 °C for 1 month.

Technical support

Scan or click the QR code for our primer Tm calculator and answers to frequently asked technical questions. For further technical support, please email technical@pcrbio.com with the following information:

- Amplicon size
- Reaction setup
- Cycling conditions
- Screen grabs of amplification traces and melting profile

TM CALCULATOR

FAQS

Product Use: Unless we agree otherwise in writing, the Goods we supply are provided:

1. For research purposes only and you should not use or rely on the Goods for diagnostic purposes. If you wish to use the Goods in a regulatory approved medical device, please contact us so that we may consider this and discuss it further with you.
2. Subject to our standard terms and conditions found at https://pcrbio.com/terms-conditions/.

Important considerations

2x VeriFi® Mix: The 2x mix contains VeriFi™ Polymerase, 6 mM $MgCl_2$, 2 mM dNTPs, enhancers and stabilizers. Further addition of PCR enhancers or $MgCl_2$to the reaction is not recommended. The buffer's composition has been optimised to maximise PCR success rates.

Template: For eukaryotic DNA use between 5 ng and 500 ng per reaction, for cDNA use below 100 ng per reaction.

Primer design: Primers should have an approximate Tm of ~60 ˚C using default Primer 3 settings (https://bioinfo.ut.ee/primer3/). The final primer concentration in the reaction should be between 0.2 µM and 0.6 µM.

Denaturation: Denaturation should be performed at 95 ˚C. However, if the presence of high GC regions results in low yields, increasing the melting temperature to 98-100 ˚C can improve the amount of product.

Annealing: To verify the best annealing temperature for your primers in our products, please visit https://pcrbio.com/resources/tm-calculator/. If troubleshooting indicates issues with primer annealing, we recommend performing a temperature gradient to experimentally determine the optimal annealing temperature. Alternatively, we recommend a 55 ˚C annealing temperature then increase in 2 ˚C increments if non-specific products are present.

Extension: Optimal extension is achieved at 72 ˚C. The optimal extension time is dependent on amplicon length and complexity of template. 30 seconds per kilobase (kb) is recommended for most applications however shorter extension times of between 10 and 30 seconds per kb are possible. Two-step cycling protocols may also be used with combined annealing and extension at 68-75 ˚C.

Fast cycling: If using faster extension times, care must be taken to prevent loading too much template DNA. If non-specific bands are visible after amplification, the amount of template DNA should be decreased.

Reaction setup

1. Prepare a master mix based on the following table:

Reagent	25 µL reaction	50 µL reaction	Final concentration	Notes
2x VeriFi® Mix	12.5 µL	25 µL	1x	
Forward primer (10 µM)	1 µL	2 µL	400 nM	See above for optimal primer design
Reverse primer (10 µM)	1 µL	2 µL	400 nM	
Template DNA	<100 ng genomic DNA <5 ng less complex DNA	<200 ng genomic DNA <10 ng less complex DNA	variable	
PCR grade dH_2O	Up to 25 µL final volume	Up to 50 µL final volume		

2. Cycle using conditions based on the following table:

Cycles	Temperature	Time	Notes
1	95 ˚C	1 min	Initial denaturation
25-35	95 ˚C 60 ˚C - 75 ˚C 72 ˚C	15 seconds 15 seconds 10 - 30 sec/kb	Denaturation Anneal Extension (*see notes above)