Date 2015.08
Version 1_03

Super-Run EX Taq DNA Polymerase

Cat No : PT-T535-250U **Size** : 250 units (50μL)
SA-T535 25units (5μL)

Concentration : 5 units/μl

Purity degree:

Nicking activity, endonuclease and exonuclease activity were not detected after the incubation of 0.6 μg of supercoiled pBR322 DNA, 0.6 μg of λ DNA or 0.6 μg of λ -Hind III digest with 10 units of this enzyme for 1 hour at 74°C.

PCR products:

As most PCR products amplified with Super-Run EX Taq that is high fidelity thermostable DNA polymerases, have one A to add at 3'-termini, the obtained PCR product can be directly used for cloning into T-vector vector. Also it is possible to clone the product in blunt-end vectors after blunting and phosphorylation of the end.

Components : ***Store at -20°C.***

- 1 tubes of 50 μl Super-Run EX Taq DNA polymerase
- 1 tube of 1ml 10X Super-Run EX Taq Buffer (Mg^{2+} plus)

General Protocol: (total 50 ul)
Set up as follows:

	Experimental Sample
Super-Run EX Taq (5 units/μl)	0.25 ul
10X Super-Run EX Taq Buffer (Mg^{2+} plus)	5 ul
dNTP Mixture (2.5mM)	4 μl (1ul for 10 mM dNTP)
Primer 1 (0.2 ~ 1.0 uM)	X μl
Primer 1 (0.2 ~ 1.0 uM)	Y μl
Template < 500 ng	Z μl
Sterilized distilled water	Up to 50 ul
Total	50 μl

- **For Experimental Sample, go to your own Cycle Conditions.**
- **When amplifying 1 kb DNA fragment: 30 cycles of 98 °C denaturation for 10 sec, 55 °C annealing for 30 sec and 72°C extension for 1 mins, Or, 30 cycles of 98 °C denaturation for 10 sec, 68°C annealing/extension for 1 min.** Denatureation condition varies depending on an used thermal cycler and tube. It is recommended for 10-30 sec at 94°C , or 1-10 sec at 98°C
- **Add 3'A-Overhangs Post-Amplification.** (This is just one method for adding 3´ adenines.)
  1. After amplifying with a Super-Run EX DNA polymerase, place vials on ice and add 0.7-1 unit of Taq polymerase per tube. Mix well. It is not necessary to change the buffer. A sufficient number of PCR products will retain the 3´ A-overhangs.
  2. Incubate the vials at 72°C for 8-10 minutes (do not cycle).
  3. clean up or gel extraction of target DNA bands before TA cloning.

For Research Using Only.
Please do not hesitate to contact us if you have any questions.

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